

International Journal of Veterinary Sciences and Animal Husbandry



ISSN: 2456-2912 VET 2024; 9(3): 252-255 © 2024 VET

www.veterinarypaper.com

Received: 01-02-2024 Accepted: 02-03-2024

G Sravani

P.G. Scholar, Department of Microbiology, N.T.R College of Veterinary and Animal Science, Gannavaram, Andhra Pradesh, India

PI Ganesan

Professor & Head, Department of Veterinary Medicine, Apollo College of Veterinary Medicine, Jaipur, Rajasthan, India

Concurrent infections of *M. audouinii* and Mucormycosis in buffaloes and attributed risk factors for fungal infections

G Sravani and PI Ganesan

Abstract

Studies on the prevalence of fungal diseases carried along with their clinical signs in an organized livestock farm in Jamdoli, Jaipur, having 19 cows, 18 Murrah buffaloes,6 Murrah calves, with varying age groups. Few Murrah milch animals and buffalo calves revealed skin lesions. Hence all the buffaloes and their calves were clinically examined and the skin scrapings were screened for their etiological agents' involved in the causation of the cutaneous lesions. In this study the adult buffaloes suffered with hair loss with white spots, alopecia, muco-cutaneous ulceration, scaling, plaques, necrotizing fasciitis and eschar. Affected animals develop discrete scaling batches of hair loss with grey-white crust that later become a thickly suppurated crust in variable locations. The lesions observed mostly on the neck, chest, face, dewlap, lower abdomen and inter-maxillary skin. Skin scrapings from this buffalo and their calves revealed concurrent fungal infections of microsporum and Mucormycosis. The risk factors attributed for the development of cutaneous lesions in these bovine populations discussed in this study.

Keywords: Buffaloes, microsporum, mucormycosis, risk factors

Introduction

Dermatophytosis is a common contagious disease caused by fungi known as dermatophytes. Most of the fungi reside in the soil and in living tissues. Zoophilic species are adapted to animals. Geophilic species live in the environment but occasionally act as parasites. In living tissues dermatophytes remain in superficial tissues such as epidermis, hair and nails. The illness causes disfiguring and uncomfortable, especially when the lesions are widespread. Economic effects, such as damage to the hides are important in livestock. In immune compromised animals the dermatophytes invade subcutaneous tissues and other sites. The pathogenic dermatophytes causing keratin digestion are trichophyton, microsporum and epidermophyton species. Most of the dermatophytes becomes adapted to people or animals and are maintained in these reservoirs. Most or all zoophilic dermatophytes are thought to be zoonotic, although some are transferred to people more often than others. There are numerous species of anthropophylic dermatophytes in the 3genera i.e trycophyton, microsporum and epidermophyton. The predominant species of dermatophytes involved in human cases vary with climate and location and other factors such as exposure to livestock, pets or exotic species. Zoophilic dermatophyte M. canis infects ruminants also. Anthropophylic dermatophytes reported infrequently in animals. Some species documented are M. audouinii, T. rubrum, T. tonsurans, T. violaceum, T. schoenleinii, and E. floccosum. Potential predisposing factors such as tumors or treatment with immunosuppressive drugs were reported in some cases. (The Centre for food security and public health, Institute for international Cooperation in Animal Biologics: Iowa state university, OIE and USDA 2013: Papini et al 2009; Nweze, 2011) [19, 20]. Fungal infections are relatively common in healthy and immunocompetent animals compared with bacterial and viral, since fungal infections are present in the environment in great numbers. (Kohler et al. 2015; Gnat et al. 2020a) [15, 13]. The prevalence of fungal infections are low and recurrent caused by true and opportunistic pathogens, noticed worldwide in animals and human beings. (Fisher et al. 2012; Bishnoi et al. 2018; Shenoy and Jayaraman 2019; Gnat et al. 2020c) [14, 9, 2, 27].

Corresponding Author: G Sravani

P.G. Scholar, Department of Microbiology, N.T.R College of Veterinary and Animal Science, Gannavaram, Andhra Pradesh, India succumbed. Gasping immediately after making a small abdominal incision might be suggestive of communication between thoracic and abdominal cavities.

Mucormycosis is an opportunistic invasive fungal infection belongs to the genus rhizopus, mucor, rhizomucor etc. (Hoffmann *et al.* 2013; Upadhyay *et al.* 2019; Walther *et al.* 2019) [14, 32, 34]. Ali R, *et al.* (2006) reported that the mucor pathogens found in soil, decaying debris and cow feed, such as straw. The natural ecological niche of mucorales fungi are soil and decaying material. Ravise *et al.* 1978 [22]; Schonmann *et al.* 1977 [24]; Connolly *et al.* 2005 [6]; Thirion-Delalande *et al.* (2005) [31] reported cutaneous mycoses due to mucor species in cattle, horses and cat.

Results and Discussion



Fig 1a &1b: Buffaloes: Combined cutaneous infection of *M. audouinii &* Mucormycosis



Fig 2: Buffaloe- M. audouinii Mucormycosis in SDA culture

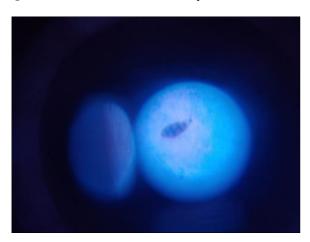


Fig 3a: Buffaloe- M. audouinii stained in LPCB



Fig 3b: M. audouinii stained in LPCB.



Fig 4a & 4b: M audouinii in rice culture-brown color

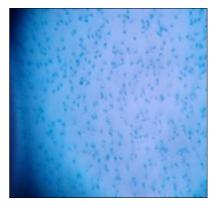


Fig 5: Mucormycosis Sporangiospores stained by LPCB

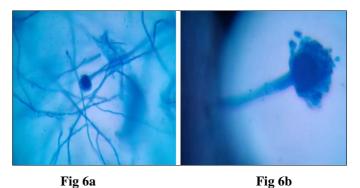


Fig (6a & 6b): Mucormycosis- Mycelium with sporongia & sporesby LPCB staining

Clinical examination of the buffaloes

In this study the buffaloes and their calves suffered with hair loss, white spots, alopecia, muco-cutaneous ulceration, scaling, plaques, necrotizing fasciitis and eschar. (Fig.1) In cattle, affected animals develop discrete scaling batches of hair loss with grey-white crust that later become a thickly

suppurated crust in variable location. (Mignon and Monod 2011; Swai and Sanka, 2012) [18, 29]. Castrejon-Perez *et al*, 2017; Lin *et al*. 2017; Sahota *et al*. (2017) [5, 23, 16] reported subcutaneous tissues and skin with subcutaneous mycoses. Ram-maert *et al*. 2014; Shields *et al*. 2019; McMahon *et al*. (2020) [21, 26, 17] reported the clinical manifestations of ecthyma-gangrenosum—like lesions, muco-cutaneous ulceration, eschar, necrotic papulo nodules, cellulytic plaques and necrotizing fasciitis or any combination of these lesions in cattle population due to Mucormycosis. In this study the buffaloes and their calves suffered with hair loss, white spots, alopecia, muco-cutaneous ulceration, scaling, plaques, necrotizing fasciitis and eschar. (Fig.1a& 1b).

Culture studies of the skin samples

The skin scrapings collected from the ailing animals subjected to culture studies using SDA revealed both microsporum species and mucormyses species (Fig2). The morphology of the culture and tests with lacto phenol cotton blue and rice grain color pigmentation studies confirmed the presence of the above fungi as M. audouinii. (Fig.3a&3b) Further studies for species wise identification of M. audouinii carried out by urease enzyme analysis and by rice culture exposure method. (Fig.4a &4b) as per the method of Ellis et al. (2013) [8] who differentiated M. canis non -sporulating strain, and M. audouinii using rice grain for its brown color pigment. Wray et al. (2008) [35] identified the mucor species by its morphophological characteristics of the filamentous growth of a vellowish/brown spreading colony. sporangiophores and spores in SDA culture. In this study also the mucor species was identified as per this procedure by its morphological structures. (Fig.5,6a & 6b).

The skin scrapings revealed the presence of *M. audouinii* and Mucormycosis by culture in Sabroud dextrose agar media (SDA) and by lactophenol cotton blue staining. Sharma *et al.* (2010) ^[25] reported the infection of cattle population with *Microsporum gypseum, Trichophyton mentagrophytes* and mucor species as a contaminant species. Sravani and Ganesan (2023) ^[28] reported the prevalence of *M. audouinii* in buffaloe calves. Vipparti 2014 ^[33]; Gawaz and Weisel (2018) ^[11] reported co-existence of yeast-like fungi and dermatophytes or mould in the same lesion.

de Crecy *et al.* 2009 ^[7]; Casadevall *et al.* (2019; Casadevall *et al.* (2020) ^[3-4] opined that the climatic changes associated with anthropogenic pressure led to adaptation of harmless fungal species to become infectious. Friedman and Schwartz (2019) ^[10] stated that the expansion of the currently known fungal pathogens happens in the areas with higher average temperatures and wetter environment. The present study was carryout in Jaipur where the city experiences both extreme weather conditions which favor the adaptations of fungal species to become infectious. These observations coincide with the outcome of the present studies.

Conclusions

Studies on the clinical examinations followed by cutaneous lesions examination were carried out in buffaloes. The buffalo population confirmed the prevalence of both microsporum and mucormyces fungal infections with cutaneous lesions. The reasons attributed for the fungal infections in cattle population are; thermal bearing capacity of the animals, anthropogenic pressure on the fungal agents to become infectious, expansion of more pathogens due to higher temperature and wetter environment, more possibilities of transmission of fungal infections from human to animals, and

the undefined source of asymptomatic & carrier animals for further infection. Studies needed to characterize the pathogens and the host - dependent disposing factors to the disease state.

References

- 1. Ali R, Khan IH. Mycotic abortion in cattle. Pak Vet J. 2006;26(1):44-6.
- 2. Bishnoi A, Vinay K, Dogra S. Emergence of recalcitrant dermatophytosis in India. Lancet Infect Dis. 2018;18:250-1.
- 3. Casadevall A. Climatic change brings the specter of new infectious diseases. J Clin Invest. 2020;130:553-5.
- 4. Casadevall A, Kontoyiannis DP, Robert V. On the emergence of *Candida auris*: climate change, azoles, swamps, and birds. mBio. 2019;10:e01397-19.
- 5. Castrejon-Perez AD, Miranda I, Welsh O, Welsh EC, Ocampo-Candiani J. Cutaneous Mucormycosis. An Bras Dermatol. 2017;92:304-11.
- 6. Connolly JH, Canfield PJ, Obendorf DL. Gross, histological and immunohistochemical features of Mucormycosis in the platypus. J Comp Pathol. 2000:123:36-46.
- 7. de Crecy E, Jarosnski S, Lyons B, Lyons T, Keyhani N. Directed evolution of a filamentous fungus for thermotolerance. BMC Biotechnol. 2009;9:74.
- 8. Ellis D. Microsporum audouinii. University of Adelaide; c2013. Available from: www.mycology.adelaide.edu.au/Fungal-Descriptions/Dermatophytes/Microsporum/Microsporum_audouinii.html
- 9. Fisher MC, Henk DA, Briggs CJ, Browstein JS, Madoff LC, McCraw SL, *et al.* Emerging fungal threats to animal, plant and ecosystem health. Nature. 2012;484:186-94.
- Friedman DZP, Schwartz IS. Emerging fungal infections: new patients, new patterns, and new pathogens. J Fungi, 2019.
- 11. Gawaz A, Weisel G. Mixed infections are a critical factor in the treatment of superficial mycoses. Mycoses. 2018;61:731-5.
- 12. Gnat S, Lagowski D, Nowakiewicz A. Major challenges and perspectives in the diagnostics and treatment of dermatophyte infections. J Appl Microbiol. 2020;129:212-32.
- 13. Gnat S, Lagowski D, Nowakiewicz A, Osinska M, Kopinski L. Population differentiation, antifungal susceptibility, and host range of Trichophyton mentagrophytes isolates causing recalcitrant infections in human and animals. Eur J Clin Microbiol Infect Dis. 2020;39:2099-113.
- 14. Hoffmann K, Pawlowska J, Walther G, Wrzosek M, de Hoog GS, Benny G, *et al.* The family structure of the Mucorales: A synoptic revision based on comprehensive multigene genealogies. Persoonia Mol Phylogeny Evol Fungi. 2013;30:57-76.
- 15. Kohler JR, Casadevall A, Perfect J. The spectrum of fungi that infects humans. Cold Spring Harb Perspect Med. 2015;5:a019273.
- 16. Lin E, Moua T, Limper AH. Pulmonary Mucormycosis: Clinical features and outcomes. Infection. 2017;45:443-8.
- 17. McMahon DE, Hysell K, Montgomery M, Frangos J. Superficial cutaneous zygomycosis presenting as resistant intertrigo: a case report. Open Forum Infect Dis; c2020. p. 7.
- 18. Mignon B, Monod M. Zoonotic infections with

- dermatophyte fungi. In: Textbook of Zoonoses. 2nd ed. New York: Oxford University Press; c2011.
- 19. Nweze EI. Dermatophytosis in domesticated animals. Rev Inst Med Trop. 2011;53(2):94-9.
- 20. Papini R, Nordoni S, Fanelli A, Mancianti F. High infection of *Trichophyton verrucosum* in calves from Central Italy. Zoonoses Public Health. 2009;56(2):59-64.
- 21. Rammaert B, Angebaut C, Scemla A, Fraitag S, Lerolle N, Lecuit M, *et al.* Mucor irregularis-associated cutaneous Mucormycosis: Case report and review. Med Mycol Case Rep. 2014;6:62-5.
- 22. Ravassi P, Fromentin H, Destombes P, Mariat F. Cerebral Mucormycosis in the cat by *Mucor pusillus*. Sabouraudia. 1978;16:291-8.
- 23. Sahota RM, Gambhir R, Anand S, Dixit A. Rhino cerebral mucormycoses: report of a rare case. Ethiop J Health Sci. 2017;27:85-90.
- 24. Schonmann M, Thomas R, Bruun U. A case of acute disseminated mucor encephalitis in a heifer. Schweiz Arch Tierheilkd. 1977;139:490-4.
- 25. Sharma DK, Joshi G, Singathia R, Lakhothia RL. Fungal infections in cattle in a Gausala at Jaipur. Haryana Vet. 2010;49:62-3.
- Shields BE, Roseenbach M, Brown-Joel Z, Berger AP, Ford BA, Want KA. Angioinvasive fungal infections impacting the skin: Background, epidemiology, and clinical presentation. J Am Acad Dermatol. 2019;80:869-80.e5.
- 27. Shenoy M, Jayaraman J. Epidemic of difficult-to-treat tinea in India: current scenario, culprits, and curbing strategies. Arch Med Health Sci. 2019;7:112.
- 28. Sravani G, Ganesan PI. International Conference on Global Perspectives in Ethno Veterinary Herbal Research for Production of Residue-Free Animal Products. VC&RI, Orathanadu, TANUVAS; c2023. p. 102-3.
- 29. Swai ES, Sanka PN. Bovine dermatophytosis caused by *Trichophyton verrucosum*; A case report. Vet Rec. 2012;5(5):297-300.
- 30. The Centre for Food Security and Public Health, Institute for International Cooperation in Animal Biologics: Iowa State University, OIE, and USDA; c2013.
- 31. Thirion-Delalande C, Guillot J, Jensen HE, Crespeau FL, Bernex F. Disseminated acute concomitant aspergillosis and mucormycoses in a pony. J Vet Med A. 2005;52:121-4.
- 32. Upadhyay V, Kumar A, Singh AK, Pandey J. Epidemiological characterization of dermatophytes at a tertiary care hospital in Eastern UP, India. Curr Med Mycol. 2019;5:1-6.
- 33. Vippartti SJ. Mixed fungal lung infection with Aspergillus fumigatus and Candida albicans in an immunocompromised patient: case report. J Clin Diagn Res; c2014. p. 8.
- 34. Walther G, Wagner L, Kurzai O. Updates on the taxonomy of Mucorales with an emphasis on clinically important taxa. J Fungi. 2019;5:106.
- 35. Wray JD, Sparkes AH, Johnson EM. Infection of the subcutis of the nose in a cat caused by Mucor species: successful treatment using posaconazole. J Feline Med Surg. 2008;10:523-7.

How to Cite This Article

Sravani G, Ganesan PI. Concurrent infections of *M. audouinii* and Mucormycosis in buffaloes and attributed risk factors for fungal infections. International Journal of Veterinary Sciences and Animal Husbandry. 2024; 9(3): 252-255.

Creative Commons (CC) License

This is an open access journal, and articles are distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 4.0 International (CC BY-NC-SA 4.0) License, which allows others to remix, tweak, and build upon the work noncommercially, as long as appropriate credit is given and the new creations are licensed under the identical terms.